

Antibacterial Activity of Graphene Oxide/g-C₃N₄ Composite through Photocatalytic Disinfection under Visible Light

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(5) Supporting Information

ABSTRACT: Carbon-based nanomaterials have been widely developed into innovative antimicrobial agents due to their advantages of high surface-to-volume ratio, extremely high mechanical strength, and distinct physicochemical properties. Here, the nanocomposite of graphene oxide/graphitic carbon nitride (GO/g-C₃N₄), a free-metal photocatalyst, was fabricated through sonication at room temperature and its antibacterial activity against *Escherichia coli* (*E. coli*) was investigated. The 100 μ g/mL GO/g-C₃N₄ composite was found to kill 97.9% of *E. coli* after 120 min visible light irradiation, which was further confirmed by fluorescent-based cell membrane integrity assay. Additionally, the holes produced by photocatalysis were confirmed by electron spin resonance (ESR) spectra and trapping experiments to participate in photocatalytic sterilization as principal active species and were further verified by transmission electron microscopy (TEM) and scanning electron microscope (SEM) to lead to the distortion and rupture of cell membrane and finally cell death. Further photoluminescence (PL) spectra, cyclic voltammetry, photocurrent generation, and impedance spectroscopy (EIS) character-



ization revealed that the introduction of GO contributed to separate photogenerated electrons and prevents the electron-hole pairs of $g-C_3N_4$ from recombing to generate more h^+ , thus directly improving the bactericidal ability of $GO/g-C_3N_4$. Reusability assays indicated that the $GO/g-C_3N_4$ retained more than 90% of activity after four cycles of use. This study facilitates an in-depth understanding of the mechanism of visible light-driven disinfection and provides an ideal candidate sterilizing agent for treating microbial-contaminated water.

KEYWORDS: GO/g-C₃N₄, Escherichia coli, Photocatalytic disinfection, Metal-free photocatalyst

INTRODUCTION

Currently, the number of people lacking access to clean water has reached nearly 900 million in the world, and about 1.5 million children under the age of five lose their lives due to the use of unsafe drinking water every year.¹ The content of Escherichia coli (E. coli) is an important indicator of clean water. Drinking water containing excessive amounts of E. coli can lead to severe inflammation and even death. Commonly used water disinfection methods include chemical and physical ones, such as the use of chlorine, chlorine dioxide, ozone, and UV irradiation. Despite effective elimination of microbial contamination, these methods have several common drawbacks. For instance, chlorination is not environmentally friendly due to its association with carcinogenic byproducts, such as trihalomethanes.² UV light disinfection is not sustainable, and when the water leaves the ultraviolet sterilization device, the surviving bacteria will breed rapidly and lead to secondary pollution. Besides, some UV-resistant E. coli strains can hardly be killed by UV light. Therefore, it is highly necessary to develop novel antimicrobial agents for effective treatment of microbialcontaminated water.

Nanoscale materials have sprung up as innovative antimicrobial agents because of their unique chemical and physical properties. Up to now, $TiO_{2^{13}}$ ZnO, MgO, Al₂O₃, AgNPs,^{4,5} and carbon-based nanomaterials⁶⁻⁸ (GO, rGO, SWCNT, C60, MWCNT) have been reported to display antibacterial activity to some extent, and among them, TiO₂ has attracted wide attention in the photocatalysis field. However, pure TiO₂ is not an ideal photocatalyst due to its limitations in the excitation wavelength of ultraviolet band, implying that it can not make full use of solar light.9 To solve this problem, TiO₂ was modified with noble metal,¹⁰ metal oxide,¹¹ and semiconductor.¹² It was reported that *E. coli* could be killed when cultured with TiO₂/Pt particles for 60-120 min under metal halide lamp irradiation.¹³ Wen et al. also reported that a new nanocomposite prepared from anatase TiO₂, Ag, and graphene showed a notable increase in both visible light absorption and photocatalytic activity.¹⁴ However, the majority of previously reported photocatalysts for bacterial inactivation contain metal nanoparticles and ions, such as $Ag^{0,15}$ Ag^+ , and Cu^{2+} , which are unfavorable for "green" water disinfection.¹⁶

Graphitic carbon nitride $(g-C_3N_4)$ is a metal-free photocatalyst consisting of tri-s-triazine subunits which are linked by planar tertiary amino groups in a layer, and it has attracted

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extensive interest among researchers due to its high photochemical stability,¹⁷ plentiful material source, inexpensive synthesis, and nontoxicity.¹⁸ These superior properties demonstrate the potential of metal-free $g-C_3N_4$ as a "sustainable" advanced photocatalyst, but pure g-C₃N₄ still has some limitations in photocatalytic efficiency because of its low electrical conductivity and rapid recombination frequency of photogenerated carriers.¹⁹ One intriguing method for improving the separation efficiency is to build heterojunction with graphene oxide (GO).^{20,21} Go has been served as a separation focus and electron acceptor to enhance the efficiency of light collection and electron-hole separation.^{22,23} When combined with g-C₃N₄, GO can rapidly transfer the photogenerated electrons to the surrounding environment, leaving little chance for the photogenerated electrons and holes to recombine.^{24,25} Recently, $GO/g-C_3N_4$ nanocompsites have been widely explored for the application as organic electrocatalysts in oxygen evolution reaction, 20,26 reduction of carbon dioxide to methane, 27,28 photoelectrochemical sensing, 29,30 fuel cell catalysis, 31,32 H₂ production, 33 and pollutant degradation.^{34,35} However, to our best knowledge, little literature is available about the function of $GO/g-C_3N_4$ as an antibacterial agent.

In this present study, we synthesized a nonmetal heterojuntion photocatalyst $GO/g-C_3N_4$ (containing 1 wt % GO) composite by sonication at room temperature and investigated its antibacterial ability on *E. coli*. Bacterial survival analyses demonstrated that $GO/g-C_3N_4$ composite could kill 97.9% of *E. coli* after 120 min visible light irradiation at the concentration of 100 $\mu g/mL$. The antibacterial activity of GO/g-C₃N₄ composite might be derived from the holes with positive charge (h⁺) produced by photocatalysis, which lead to the distortion and rupture of cell membrane (Scheme 1).

EXPERIMENTAL SECTION

Chemicals. In this work, graphite powder, urea, NaCl, yeast extract, tryptone, Cr (VI), and sodium oxalate were of analytical grade and utilized as purchased without further purification. Chemicals were purchased from Aladdin Corporation (Shanghai, China). The *E. coli*

Scheme 1. Possible Mechanisms of the Antibacterial Activity of $GO/g-C_3N_4$ Composite



strains were maintained by the State Key Laboratory of Agricultural Microbiology of Huazhong Agricultural University.

Synthesis of GO. GO was synthesized from natural graphite powders according to the typical Hummers method with minor modification.³⁶ Briefly, 1.0 g of nature graphite power was mixed with 23 mL sulfuric acid (95%) under ice bath, followed by addition of 3.0 g KMnO₄ under vigorous stirring to ensure thorough dissolution. After adequate oxidation, the mixture was ultrasonicated at room temperature for 10 h. Next, the mixture was quickly transferred into 46 mL H_2O and 150 mL H_2O_2 solution (9%). Finally, the solution was filtered, washed with 0.2 M HCl, and freeze-dried for 12 h for further use.

Synthesis of $g-C_3N_4$ **.** The $g-C_3N_4$ was prepared by heating urea directly.³⁷ Three grams of urea powder were heated to 600 °C at a rate of 5 °C per minute. After 4 h of heating, the urea sample was cooled down to room temperature and then ground to powder.

Fabrication of GO/g-C₃N₄ Composite. The GO/g-C₃N₄ composite photocatalyst was prepared by a straightforward sonochemical method. First, 100 mg g-C₃N₄ was suspended in 80 mL ultrapure water while 1.0 mg GO was mixed with 20 mL ultrapure water. Then, the two solutions were ultrasonicated separately for 1 h. Next, the GO and g-C₃N₄ dispersions were mixed and ultrasonicated for 2 h. After freeze-drying for 24 h, the product was obtained.

Characterization of GO/g-C₃N₄ Composite. The morphology of the as-prepared photocatalysts was characterized via transmission electron microscopy (TEM), with the images obtained through a Hitachi H-7650 EM operated at 300 kV. The crystal structures were measured by X-ray diffractometer (XRD) using Cu–K α radiation. The surface functional groups of obtained samples were investigated via Fourier transform infrared spectroscopy (FT-IR, Nicolet Avatar-330) using the KBr pellet pressing method. Synergetic effects of GO and g-C₃N₄ on light absorption properties were studied using UV-vis diffuse reflectance spectroscopy (DRS, Cary 5000 UV-vis-NIR) with an integrating sphere using BaSO_4 as the blank reference. The photoluminescence (PL, Hitachi F-4600) spectral analysis was performed to probe the effect of the GO modification. Photocurrent, cyclic voltammograms and electrochemical impedance spectral (EIS) tests were performed by a CHI 660D electrochemical workstation (Chenhua Instrument, Shanghai, China), using Pt foil as a counter electrode, Ag/AgCl (saturated KCl) as a reference electrode, and 0.1 M Na2SO4 solution as the electrolyte. X-ray photoelectron spectra (XPS) were measured using a Thermo VG Multilb 2000 spectrometer with a homochromous Al–K α radiation source.

Antibacterial Activity Test. *E. coli* was applied to assess the antibacterial effect of $GO/g-C_3N_4$ composite. Briefly, 3.0 μ L of *E. coli* was added into 3.0 mL LB liquid medium, which was then transferred into a 37 °C incubator with gentle shaking. After 16 h incubation, the cell count approximated 10⁹ colony forming units (CFU) per milliliter. Then bacterial cells were collected by centrifugation at 5000 rpm for 10 min, followed by washing three times to remove the supernatant, and then resuspended in PBS buffer (pH = 7.4). The bacterial suspensions were diluted to 10⁷ CFU/mL.

Next, 100 μ L composite photocatalyst suspensions with different concentrations were added to 3.0 mL of bacterial solutions to ensure that the ultimate concentrations of the GO/g-C₃N₄ composite were 25, 50, and 100 μ g/mL. Then the mixture was treated for 2 h under a 300 W xenon lamp mounted with UV filter ($\lambda < 420$ nm). Meanwhile, the light intensity was maintained at 300 mW/cm⁻¹ throughout the experiments. The antibacterial experiment was conducted at room temperature by constantly stirring the mixture at 500 rpm and collecting 100 μ L sample from the mixture per 30 min. After dilution to a suitable concentration with PBS buffer, 100 μ L diluted sample was quickly spread on an LB agar culture plate and cultured at 37 °C for 17 h. The survival rate was estimated by the plate count method. The blank control group without any GO/g-C₃N₄ composite and the light control without any irradiation were also prepared. All the experiments were performed in triplicate.

Morphological Analysis of Bacterial Cells. SEM and TEM were often applied to characterize the morphology of *E. coli* untreated and treated with $GO/g-C_3N_4$ composite under visible light or dark. The



Figure 1. TEM pictures of (a) g-C₃N₄, (b) GO/g-C₃N₄. (c) XRD and (d) FT-IR spectra of GO, g-C₃N₄, and GO/g-C₃N₄.



Figure 2. XPS spectra of GO/g-C₃N₄: (a) survey spectrum, (b) C 1s spectrum, (c) N 1s spectrum, and (d) O 1s spectrum.

bacterial suspensions were exposed to $GO/g-C_3N_4$ under irradiation for 2 h, then centrifugated at 8000 rpm for 5 min. Next, the condensed cells were fixed with cold 2.5% glutaraldehyde for 1 h, washed twice with PBS buffer, dehydrated successively with 30, 50, 70, 90, and 100% ethanol for 15 min and then freeze-dried. Finally, the obtained samples were observed separately under a SEM (JEOL JSM-6700F) and TEM (HITACHI H-7650).

Fluorescent-Based Cell Membrane Integrity Analysis. *E. coli* cells in logarithmic phase were cultured in LB medium, followed by mock-treatment or treatment with $100 \ \mu g/mL \ GO/g-C_3N_4$ composite under visible light for 2 h. The cells were collected by centrifugating, followed by staining with propidium iodide (PI, $10 \ \mu g/mL$) and 4'-6-

diamidino-2-phenylindole (DAPI, 5 μ g/mL) for 15 and 5 min, respectively. After washing twice to discard the excess dye, the bacterial cells were observed with a confocal microscope (OLYMPUS FV1000).

Cytotoxicity Test. Five ×10³ cells/well Hela cells in 96-well plates were cultured until 80–90% confluence, followed by supplementation of different concentrations (5, 10, 20, 50, 100, and 200 μ g/mL) of GO/g-C₃N₄. After 24 h incubation, each well was supplemented with 20 μ L MTT reagent (3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2-*H*-tetrazolium bromide) and cultured for another 4 h. Then the supernatant was discarded, and the formazan was dissolved in 150 μ L dimethyl sulfoxide (DMSO). Finally, absorbance at 490 nm was



Figure 3. (a) UV-vis diffuse reflectance images of pure $g-C_3N_4$, GO, and $GO/g-C_3N_4$ nanocomposites. (b) Plots of converted Kubelka–Munk functions versus light energy.

determined by an enzyme linked immunosorbent assay (ELISA) microplate reader. $^{\rm 38}$

RESULTS AND DISCUSSION

Characterization of GO/g-C₃N₄. Figure 1 shows the morphology and microstructure of the $GO/g-C_3N_4$ (containing 1 wt % GO) composite characterized by TEM, XRD, and FT-IR. As displayed in Figure 1a and b, g-C₃N₄ was well spread and attached to GO, and its size was in the range of tens to hundreds of nanometers. In the XRD of GO, $g-C_3N_4$ and GO/ $g-C_3N_4$ (Figure 1c), the peak at 10.30° of GO was attributed to the interlayer spacing (001) diffraction peak, with an interlayer spacing distance of 0.86 nm.^{39,40} The $g-C_3N_4$ had two intensive peaks at 13.12° and 27.50°. The former represented the (100) diffraction peak of in-plane structural packing motif of tristriazine units, and the packing distance was 0.671 nm, while the latter represented an interlayer stacking (002) diffraction peak of aromatic segments, and the stacking distance was 0.324 nm.⁴¹ There was no obvious difference between g-C₃N₄ and $GO/g-C_3N_4$ in scanning spectra, suggesting no effect of the modification with 1% GO on the g-C₃N₄ lattice structure.⁴²

The surface functional groups of GO, g-C₃N₄, and GO/g- C_3N_4 were measured by the Fourier transform infrared (FTIR) spectroscopy. In Figure 1d, the wider band at 3250–3750 cm⁻¹ of GO was attributed to the stretching vibrations of -OH and the physically absorbed H₂O; the high-intensity peaks around 1735 and 1612 cm⁻¹ to the C=O stretching vibration and distortion vibration of intercalated water; the bands at 1414, 1224, and 1046 cm^{-1} to the C–O, C–OH, and C–O–C bonds, respectively; the peak around 789 cm⁻¹ to the absorption of epoxide group;⁴³ the peaks at 1637, 1571, 1404, 1318, and 1238 cm⁻¹ of pure $g-C_3N_4$ to the characteristic stretching vibrations of C-N heterocycles; and the peak at 814 cm⁻¹ to the typical stretching vibration of triazine units.⁴⁴ For the $GO/g-C_3N_4$ composite, due to the low GO content in the composite, the representative peaks of CN heterocycles could be only observed in the spectrum of $g-C_3N_4$.

The chemical structure of $GO/g-C_3N_4$ was examined by X-ray photoelectron spectroscopy (XPS). In Figure 2b, the notably unsymmetrical and obvious C 1s peaks showed the coexistence of diacritical matrix, and the C 1s peaks at 284.7, 285.2, 286.4, 288.1, and 288.5 eV could represent sp²-hybridized carbons (C–C), C–OH, C–N–C, C=O–OH, and C–(N)₃ bonds, respectively, through signal deconvolution via Gaussian curve fitting.^{45–47} The N 1s peak at 398.5 eV was assigned to the sp² N contained in triazine rings, and the peaks at 399.8 and 401 eV were attributed to the N–(C)₃ and –NH₂ or =NH groups (Figure 2c).⁴⁸ The O 1s peaks at 532 and 533

eV were associated with O-C-N and O=C bonds, while the peak at 540 eV was due to the adsorbed O_2 (Figure 2d).⁴⁹

Optical Property. Optical properties and band positions of samples were investigated using UV-vis diffuse reflectance spectroscopy and Mott-Schottky plots. In Figure 3a, the band gap absorption edge of both $g-C_3N_4$ and $GO/g-C_3N_4$ was around 460 nm, suggesting carbon was not contained in the g- C_3N_4 lattice and that the GO sheet was only used as a matrix to fix $g-C_3N_4$ nanosheets.²² In addition, $GO/g-C_3N_4$ showed strong absorption over the whole range of the investigated wavelengths, which resulted from the strong absorption of GO in visible light region.²⁴ Besides, the band gap energies were estimated to be 2.73 and 2.51 eV for $g-C_3N_4$ and $GO/g-C_3N_4$ (Figure 3b). The narrowing band gap and enhanced visible light utilization of the nanocomposites can be ascribed to graphene hybridization, which could result in the electronic transition between $g-C_3N_4$ and graphene. rGO modification in g-C₃N₄ has been reported to improve light utilization and thus photocatalytic efficiency.⁵⁰ Ong et al. also showed that the exceptional enhancement of light absorption and the extension of absorption edge might result from the effect of graphene on the hybrid nanostructures.²⁷ Our results were similar to the previous reports,^{27,50} suggesting that GO enhances the light aggregating efficiency of the $GO/g-C_3N_4$ composite.

Antibacterial Activity of $GO/g-C_3N_4$ Composite. *E. coli* was used as a model microorganism to estimate the photocatalytic disinfection ability of $GO/g-C_3N_4$. As shown in Figure 4, in blank control (under light irradiation without catalysts), there were no bacteria inactivated, suggesting that bright light had no effect on the bacteria. Meanwhile, in the dark treatment (catalysts without light irradiation), no bacterial disinfection was detected, demonstrating the nontoxicity of the



Figure 4. Photocatalytic disinfection ability of GO, g- C_3N_4 , and GO/g- C_3N_4 . Results were reported as the mean \pm SD from three independent experiments.

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photocatalyst to *E. coli*. In Figure 4, it can also be seen that 100 μ g/mL GO had weak disinfection ability, which was consistent with the results of previous literature.⁵¹ At the same concentration, GO/g-C₃N₄ composite showed significantly higher disinfection efficiency than the GO and g-C₃N₄ in killing *E. coli*. After 120 min treatment with GO/g-C₃N₄ composite, the survival rate of *E. coli* was only 2.1%, and the antibacterial ability of GO/g-C₃N₄ against *E. coli* was also dose-dependent.

The reliability of the CFU method in the above test was verified by a fluorescent-based cell live/dead test using the confocal laser scanning fluorescence microscopy imaging system. Bacterial suspension was stained by DAPI and PI fluorescence dye reagent. DAPI can quickly pass through the unbroken membrane of live cells and combine with DNA in nucleus, while PI can only pass through the broken membrane and combine with double strand DNA. In Figure 5a, few dead



Figure 5. Confocal fluorescent assays of live and dead *E. coli* cells in the absence (a and b) presence of 100 μ g/mL GO/g-C₃N₄ under irradiation for 2 h.

cells could be observed in the untreated groups, while in Figure 5b, almost all the *E. coli* cells treated with 100 μ g/mL GO/g-C₃N₄ under irradiation for 2 h were stained by PI, suggesting that most *E. coli* cells died due to the rupture of cell membrane.

E. coli Structure Observation by SEM and TEM after Treatment with GO/g-C₃N₄ Nanocomposite. Direct interactions between biological cells and nanomaterials are usually evaluated using scanning electron microscopy (SEM) and transmission electron microscopy (TEM).⁴⁰ To observe the changes in the structure of the treated E. coli, SEM imaging was performed. In Figure 6a, the control E. coli cells were representatively rod-shaped with smooth and intact cell membrane. There were also no obvious changes detected in the morphology of E. coli treated with visible light but without $GO/g-C_3N_4$ composite or treated with $GO/g-C_3N_4$ in the dark (Figure 6b and c). However, after treatment with $GO/g-C_3N_4$ under visible light for 120 min, the cell wall became wrinkled, the shape of the cell was completely deformed, and the cell membrane was obviously damaged. In addition, the size of the cells was very uneven, with some cells almost twice longer than others (Figure 6d), and cell elongation is a classical bacterial response to stresses including exposure to biocides. These observations indicated that the visible light (without GO/g- C_3N_4 composite) or $GO/g-C_3N_4$ composite (in the dark condition) has no effect on E. coli cells, but some active species

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Figure 6. SEM pictures of *E. coli* (a) untreated or treated with (b) visible light but without $GO/g-C_3N_4$ composite, (c) $100 \ \mu g/mL \ GO/g-C_3N_4$ composite in the dark, and (d) $100 \ \mu g/mL \ GO/g-C_3N_4$ composite under light irradiation for 2 h. Scare bars: $1 \ \mu m$.

are generated by $GO/g-C_3N_4$ composite under light irradiation, such as h^+ , $\bullet O_2^-$, and H_2O_2 , which interact with the cell membrane and eventually lead to cell death.

The interactions between the *E. coli* and $GO/g-C_3N_4$ were further confirmed by TEM imaging. In Figure 7a–c, the *E. coli*



Figure 7. TEM pictures of *E. coli* (a) untreated or treated with (b) visible light but without GO/g-C₃N₄ composite, (c) 100 μ g/mL GO/g-C₃N₄ composite in the dark, and (d) 100 μ g/mL GO/g-C₃N₄ composite under light irradiation for 2 h. Scale bars: 0.5 μ m.

cells untreated, treated with visible light but without GO/g-C₃N₄ composite, or treated with 100 μ g/mL GO/g-C₃N₄ in the dark showed the typical rod-shape morphology with an intact structure. However, after 2 h of treatment with 100 μ g/mL GO/g-C₃N₄ under irradiation, the bacterial cells were deformed and collapsed and the regular cell structure was destroyed (Figure 7d). These observations were consistent with those of SEM imaging.

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Figure 8. ESR images of (a) DMPO- \bullet OH and (b) DMPO- \bullet O₂⁻ for g-C₃N₄ and GO/g-C₃N₄ under visible light irradiation.

Cytotoxicity. A good cell compatibility is essential for the use of a composite material as a water disinfectant. The effect of $GO/g-C_3N_4$ on the viability of Hela cell was evaluated using an MTT cytotoxicity assay. When compared with the control, $GO/g-C_3N_4$ showed no obvious influence on the proliferation of Hela cell in the 5–100 μ g/mL concentration range (Figure S1). Even at the high concentration of 100 μ g/mL, the GO/g-C₃N₄ showed little toxicity to cells and the cell viability was still 88%. At 200 μ g/mL GO/g-C₃N₄, the cell viability was close to 80%, which was similar to the result of a previous study.⁵² In addition, GO/g-C₃N₄ was insoluble in water, it would quickly precipitate at the end of the sterilization experiment, with the residual GO/g-C₃N₄ concentration in the supernatant much lower than the initial $GO/g-C_3N_4$ concentration. The effect of residual GO/g-C₃N₄ on cell viability was evaluated by centrifuging the supernatant at 10 000 rpm for 15 min, then dissolving the precipitation in cell culture media and using the mixture for cytotoxicity test. As shown in Figure S1, the residual GO/g-C₃N₄ in the supernatant had little effect on cell viability (97%).

Mechanism of Photocatalytic Disinfection. The h⁺, $\bullet O_2^-$, $\bullet OH$, and H_2O_2 produced in the photocatalytic process are generally considered to be the active species for disinfection. Thus, the hydroxyl radical ($\bullet OH$) and superoxide radical anion ($\bullet O_2^-$) generated by g-C₃N₄ and GO/g-C₃N₄ under visible light irradiation were detected using ESR spin-trap (DMPO) method (Figure 8). As expected, GO/g-C₃N₄ photogenerated more intense typical DMPO- $\bullet O_2^-$ and DMPO- $\bullet OH$ signals than g-C₃N₄, indicating potential participation of $\bullet O_2^-$ and $\bullet OH$ in photocatalytic disinfection. Since the valence band potential of g-C₃N₄ (1.57 V vs NHE) was deficient for oxidation of water or surface hydroxyl group (2.38 V vs NHE) to generate $\bullet OH$, the strong DMPO- $\bullet OH$ signals implied the further conversion of $\bullet O_2^-$ (Figure 8b).⁵³

The dominance of active species during photocatalytic disinfection was examined by employing isopropanol, $K_2Cr_2O_7$, superoxide dismutase (SOD) and soduim oxalate to scavenge hydroxyl radicals (\bullet OH), electron (e⁻), superoxide radical anions ($\bullet O_2^-$), and hole (h⁺), respectively (Figure 9).⁵⁴ With the addition of isopropanol, $K_2Cr_2O_7$ and SOD, only slight changes were detected in the antibacterial ability, suggesting that \bullet OH, e⁻, and $\bullet O_2^-$ had no effect on the antibacterial process. However, when sodium oxalate was added, the antibacterial ability was decreased obviously, with a bacterial survival rate of ~60% after 120 min irradiation, indicating that h⁺ served as the main active species for photocatalytic sterilization.

Mechanism of Enhanced Photocatalytic Antibacterial Activity. To explore the photocatalytic antibacterial mecha-



Figure 9. Photocatalytic disinfection ability of GO/g-C₃N₄ with different scavengers (*i*-PrOH $\rightarrow \bullet$ OH, K₂Cr₂O₇ $\rightarrow e^-$, SOD $\rightarrow \bullet$ O₂⁻, soduim oxalate $\rightarrow h^+$). Results were reported as the mean \pm SD from three independent experiments.

nism of g-C₃N₄ and GO/g-C₃N₄, the separation efficiency was studied by photoluminescence (PL) spectroscopy. In Figure 10a, the emission peaks of pure g-C₃N₄ and GO/g-C₃N₄ were around 450 nm, corresponding to the band gap of g-C₃N₄ (2.7 eV).⁵⁵ The peak of pure g-C₃N₄ was obviously higher than that of GO/g-C₃N₄, indicating that addition of GO effectively prevented the photogenerated electron—hole pairs from recombination.^{24,50} Meanwhile, the CB minimum of g-C₃N₄ was -1.73 eV vs NHE as measured by the cyclic voltammetry method (Figure 10b). These results suggest that the successful electron transfer was due to the reason that Fermi level of GO (-0.08 eV vs NHE) was lower than the CB of g-C₃N₄ and GO, which prevents direct recombination of electrons and holes.²⁸

Photocurrent generation and electrochemical impedance spectroscopy were used to examine the electron generation and charge transfer characteristics of $g-C_3N_4$, GO and GO/g- C_3N_4 . Under visible light (Figure 10c), the photocurrent response of pure $g-C_3N_4$ was notably lower than that of GO/g- C_3N_4 , which was concordant with the result of PL. In Figure 10d, the arc radius on the EIS Nyquist diagram of GO/g- C_3N_4 composites was less than that of pure $g-C_3N_4$. These results suggested that, under visible light irradiation, GO/g- C_3N_4 , showed better charge transport characteristics than $g-C_3N_4$, which may facilitate charge transfer.

Stability of GO/g-C₃N₄ Photocatalyst. A vital indicator for the practical application of a photocatalyst is stability. In this study, the stability of GO/g-C₃N₄ for the photocatalytic disinfection of *E. coli* under visible light was tested by four cycles of use of the photocatalyst under the same condition. Specifically, the photocatalyst was recollected by centrifuging and the new bacterial suspension was used in each cycle. As



Figure 10. Photoluminescence spectra (a), cyclic voltammograms (b), photocurrent responses (c), and electrochemical impedance spectra (d) of g- C_3N_4 , GO and GO/g- C_3N_4 in 0.1 M Na₂SO₄ aqueous solution under visible light irradiation.

shown in Figure 11, the photocatalyst exhibited stable and high photocatalytic disinfection performance in the four cycles.



Figure 11. Repeated experiments of photocatalytic disinfection of 100 μ g/mL GO/g-C₃N₄ under visible light.

CONCLUSIONS

In this study, $GO/g-C_3N_4$ composite (containing 1 wt % GO) was successfully synthesized by a facile sonochemical method, and analyzed by standard spectroscopic techniques (UV-vis diffuse reflectance, photoluminescence, XRD, and FTIR spectrosopy) as well as XPS and TEM. The uniform distribution of GO sheets in the layered g-C₃N₄ structure suggest a negligible influence of GO modification on the g-C₃N₄ lattice structure. This is probably the first report to demonstrate that GO/g-C₃N₄ has superior antibacterial activity against E.coli. At 100 μ g/mL, the GO/g-C₃N₄ composite could kill 97.9% of E. coli after 120 min visible light irradiation. The holes (h⁺) produced by photocatalysis were confirmed to be the major active species for photocatalytic sterilization and might cause the distortion and rupture of cell membrane. Further analyses revealed that the introduction of GO not only reinforced light absorption and photocurrent but also largely improved the photogenerated electron-hole separation, leading to an increase of the number of holes. Furthermore,

 $\rm GO/g-C_3N_4$ exhibited stable and high photocatalytic disinfection efficiency even after four cycles of use. These encouraging results provide experimental basis for further development of $\rm GO/g-C_3N_4$ composite as an ideal photocatalyst for water sterilization.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acssuschemeng.7b01431.

MTT results for Hela cells exposed to various concentrations of $GO/g-C_3N_4$ (PDF)

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Notes

The authors declare no competing financial interest.

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